

We can produce and scale-up mammalian phase I and II metabolites using microbial catalysts, mammalian tissue fractions and recombinant enzymes:

- For DMPK / ADME / TOX
- For Met ID
- As standards for quantitation
- For bioactivity testing
- For stability studies

Proven Reactions

- Methyl hydroxylation
- Methylene hydroxylation
- Methine hydroxylation
- Aromatic hydroxylation
- N-oxidation
- N-methylation
- N-dealkylation
- N-acetylation
- O-dealkylation
- Carbonyl reduction
- Heterocycle oxidation via aldehyde oxidase
- Aromatic O-glucuronidation
- Aromatic N-glucuronidation
- Non-aromatic O-glucuronidation
- Non-aromatic N-glucuronidation
- Acyl-glucuronidation
- Other glycosidations (AgChem)
- N-sulfation
- O-sulfation
- Thiol conjugation (GSH/NAC)
- Transamination
- Amino acid conjugations
- Sequential reactions e.g. hydroxylation & glucuronidation

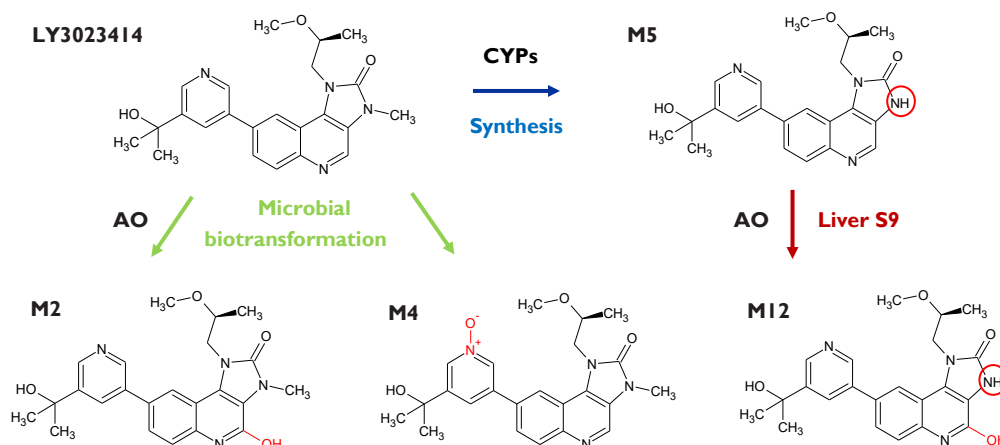
For more information contact us at mail@hyphadiscovery.com

ABOUT HYPHA DISCOVERY

Hypha Discovery Ltd is a UK-based microbial biotechnology company providing solutions to pharmaceutical and agrochemical R&D partners through the production of mammalian and microbial metabolites, as well as specialising in microbially-derived chemicals. We have an extensive client base and work with many of the top pharma and agrochemical companies worldwide.

Identifying and scaling up AO metabolites

Provision of multiple human metabolites via biotransformation



Provision of human CYP and non-CYP metabolites M2, M4 and M12 at multi mg scale utilising both microbial biotransformation and liver S9 incubations.

There has been a notable increase in metabolism of new drug candidates through non-CYP phase I pathways such as those mediated via aldehyde oxidase (AO).¹ Further, mixed AO/P450 substrates may be subject to metabolic shunting, an important consideration during toxicology and DDI assessment of these drugs.² Access to metabolites may thus be important to consider for drugs with mixed metabolism.

Zhou *et al.* presented a poster at the 2018 ISSX meeting in Montreal on "Elimination of [¹⁴C]-LY3023414 by Aldehyde Oxidase and CYP Enzymes in Humans Following Oral Administration." Both AO and CYP enzymes were responsible for the metabolic clearance of LY3023414 with the non-CYP enzymes mediating approximately half of the clearance of the drug. The predominant metabolic clearance pathways were aromatic hydroxylation of the quinoline moiety (M2), N-demethylation (M5) and quinoline oxidation with N-demethylation (M12).

No metabolism was observed when tested vs 5 human recombinant CYPs, however screening of LY3023414 against a subset of Hypha's microbial biotransforming strains generated a number of metabolites. The best strain was scaled-up to 6L to access target metabolites M2 and M4. Subsequent incubation of the synthesised intermediate M5 vs Cyno S9 enabled production of a further target metabolite M12. Metabolites were purified to > 95% purity by Hypha and the structures confirmed by LC-MS and NMR. The **AO mediated hydroxylated metabolite (M2, 20.1mg)** and an **N-oxide (M4, 66.3mg)** were made via microbial biosynthesis and a **CYP/AO mediated metabolite (M12, 18.4mg)** was generated through liver S9 incubations.

Use of combined biotransformation approaches can therefore enable access to metabolites created through different pathways, including AO and CYP mediated metabolites.

¹Rashidi & Soltani, 2017. Expert Opin. Drug Discov.12 (3), 305-316.

²Crouch *et al.*, 2016. Drug Metab Dispos 44, 1296-1303.